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Synthesis of Phosphoramidate Analogues of Sphinganine-1-phosphate

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Abstract: Phosphoramidate derivatives **13** and **15** were prepared as analogues of sphinganine-1-phosphate (**4**). Key steps of the synthesis are the alkylation of N-methoxy-N-methylcarboxamide of the diamino-propanoic acid **7** and the subsequent reduction of the intermediately formed ketone **8** affording the diastereomeric alcohols **9**. Deprotection and phosphorylation of **9** lead to the desired products **13** and **15**.

Sphingosine **1** and sphinganine **2** (fig. 1) are the long-chain bases most abundant in cellular sphingolipids, e. g. ceramide, sphingomyelin, cerebrosides and gangliosides.^{1,2} Sphinganine **2** itself is an intermediate in the biosynthesis of sphingolipids whereas sphingosine **1** is generated as a catabolic intermediate in the degradation of ceramide.³ Studies on the biological function of sphingosine and related structures showed that these molecules are reversible inhibitors of protein kinase C.⁴ They may act as endogenous modulators of cell function and possibly as second messengers.⁵ Recently, it was demonstrated that low concentrations of sphingosine increase proliferation of quiescent Swiss 3T3 fibroblasts acting in a fundamentally different, protein kinase C-independent pathway.⁶

Sphingosine-1-phosphate **3** and sphinganine-1-phosphate **4** are the initial intermediates in the catabolism of the long-chain sphingoid bases. They are formed by the cytosolic enzyme sphingosine kinase^{7,8} which

catalyses the ATP dependent phosphorylation step at the 1-OH-position (fig. 1). The 1-phosphates **3** and **4** are cleaved by the action of a pyridoxal-phosphate dependent lyase to yield a fatty aldehyde and ethanolamine phosphate.⁹ Another primary metabolic product of sphingosine-1-phosphate in cultured skin fibroblasts appeared to be sphingosine, indicating the action of a phosphatase.¹⁰ Comparison of the rates of cleavage by the action of the sphingosine-1-phosphate lyase with those of dephosphorylation indicates that dephosphorylation is at least as active as cleavage. The fast turnover of sphingosine-1-phosphate certainly favours a second-messenger role of this lipid intermediate.

The phosphorylated sphingoid bases are not only intermediary catabolites but also bioactive lipids with important functions including elevation of phosphatidic acid levels¹¹ and activation of the DNA binding activity of AP-1.¹² Sphingosine-1-phosphate strongly mimics PDGF (platelet-derived growth factor)-receptor induced chemotactic signal transduction favoring actin filament disassembly. This excessive and prolonged signaling results in a marked inhibition of cell spreading, of extension of the leading lamellae toward PDGF, and of chemotaxis toward PDGF.¹³

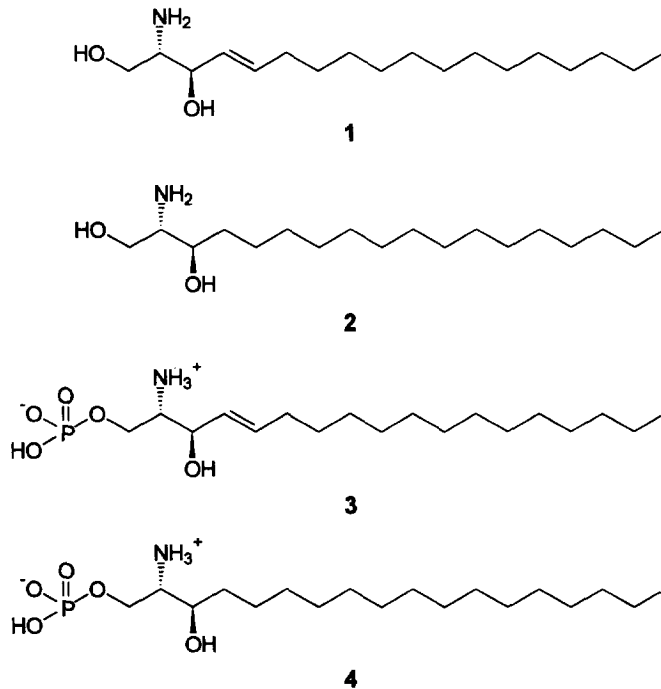


Figure 1

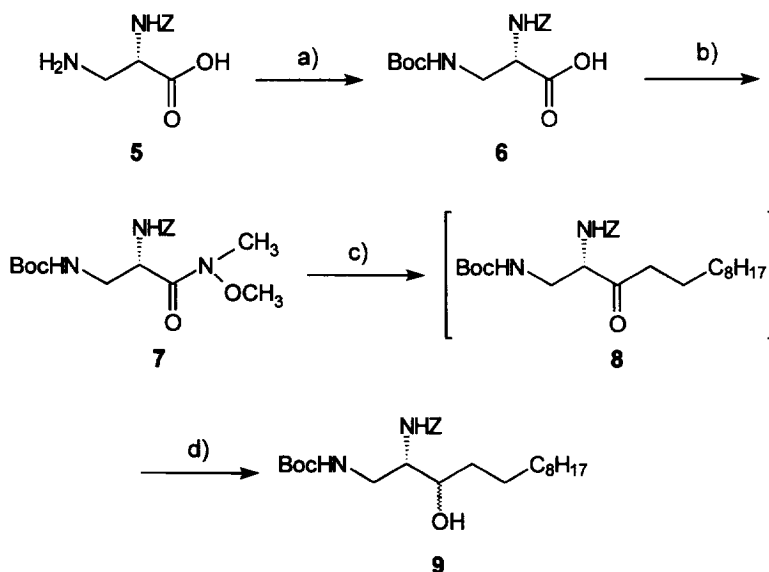
Furthermore, mitogenic concentrations of sphingosine-1-phosphate stimulate production of inositol phosphates, which can be inhibited by pertussis toxin, while the response to bradykinin is not effected. Sphingosine-1-phosphate decreases cellular cAMP levels and also causes a drastic decrease in isoproterenol- and forskolin-stimulated cAMP accumulation. These results suggest that some of the sphingosine-1-phosphate-induced signaling pathways are mediated by G proteins that are substrates for pertussis toxin.¹⁴

Sphingosine-1-phosphate induces a transient increase in intracellular free calcium independent of extracellular calcium concentrations.^{15,16} It was demonstrated that sphingosine-1-phosphate inhibits motility of melanoma cells in very low concentrations (10-100 nM), in which sphingosine shows no inhibitory effect.^{17,18} These tools suggest that sphingosine-1-phosphate may have antimetastatic and inflammatory properties.

Therefore, these findings propose that sphingosine-1-phosphate (**3**) and sphinganine-1-phosphate (**4**) have appropriate properties to function as intracellular lipid second messengers that are involved in calcium release and the regulation of the cell growth induced by sphingosine (**1**).

In this paper we describe the synthesis of **13** and **15** as structural analogues of sphinganine-1-phosphate (**4**) modified in the headgroup whereas sphingosine-1-phosphate (**3**) and sphinganine-1-phosphate (**4**) are subjected to fast turnover. **13** and **15** are metabolically resistant. Therefore, they may be suitable tools to study the intracellular functions of sphingosine-1-phosphate (**3**) and sphingosine-1-phosphate (**4**). Compared to natural sphinganine-1-phosphate (**4**), the P-O bond of the phosphoric acid ester in the 1-position of the sphingoid backbone is substituted by a P-N bond. Phosphorylated amines are close analogues of the isosteric phosphate esters with respect to geometry, bond lengths and hydrogen bonding capacity.¹⁹ Therefore, a number of phosphorylated amino analogues of metabolic intermediates have been synthesized.²⁰ The major drawback of the phosphoramides and phosphoramidates consists in their acid lability. Their resistance towards enzymes transferring phosphoryl groups is varying and depends on the enzyme.¹⁹

In order to clarify and investigate the biological functions of the metabolically labile sphingosine-1-phosphate and sphinganine-1-phosphate and in addition to the synthesis of the corresponding phosphonates analogues²¹, we synthesized the phosphoramides **13** and **15**.

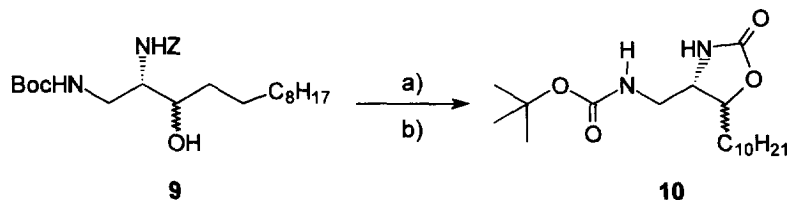


Scheme 1: a) Boc₂O, dioxane/water, 80%; b) EtOCOCl, THF, N, O-dimethyl hydroxylamine hydrochloride, THF, 62%; c) decylmagnesium bromide, THF; d) sodium borohydride, 2-propanol, 48%.

The synthesis started with protection of the N_α -benzyloxycarbonyl-L-2,3-diaminopropanoic acid (**5**) with tert-butyl dicarbonate and triethylamine in dioxane/water following previous reports²² (scheme 1).

Treatment of the N_α -benzyloxycarbonyl- N_β -tert-butoxycarbonyl-L-2,3-diaminopropanoic acid (**6**) with N,O-dimethylhydroxylamine hydrochloride in the presence of ethyl chloroformate and triethylamine according to König and Geiger²³ led to the amide **7**. Alkylation with decylmagnesium bromide following the procedure previously reported^{24,25} yielded ketone **8** that was reduced without further purification by sodium borohydride^{26,27,28} forming the diastereomeric amino alcohols **9** as 3 : 1 mixture.

The relative configuration of the alcohols **9** was established after transformation into the corresponding oxazolidinones **10**^{29,30} (scheme 2). Generally, the coupling constants of cis-isomers (*erythro*) of such 4,5-disubstituted oxazolidinones are greater than those of the trans-isomer (*threo*).³¹ According to ¹H NMR investigations of **10**, the major diastereomer is *erythro*-configured.

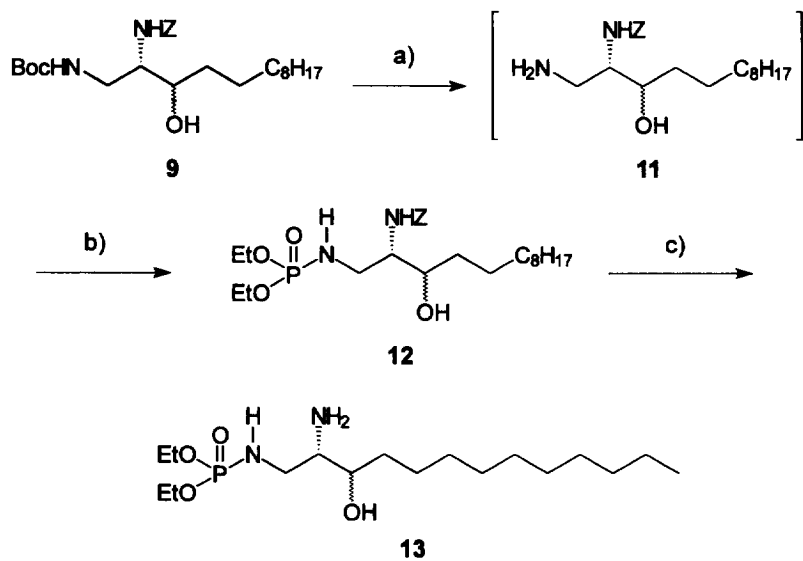


Scheme 2: a) H_2 , Pd/C, MeOH, 95%, b) $(Cl_3CO)_2CO$, dichloromethane, triethylamine, 67%

The enantiomeric purity of the alcohols **9** was assured by the Mosher method.³² The ¹H NMR spectrum of the corresponding Mosher derivatives of **9** is consistent with a racemization degree minor than 10%.

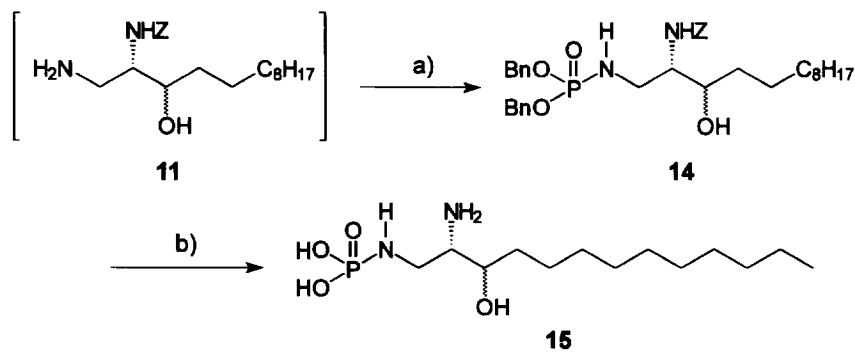
Removal of the Boc group with HCl gas in methanol and subsequent treatment with saturated sodium hydrogencarbonate solution afforded the crude amines **11** (scheme 3). The amines **11** were converted without further purification into the corresponding phosphoramides **12** by treatment with diethylchloro phosphite in chloroform^{33,34} (scheme 3).

Removal of the Z group of compounds **12** was achieved by hydrogenation with palladium on charcoal and afforded compound **13** in good overall yield. All attempts to hydrolyze the phosphoramidate diethylesters of **13** with HCl led to the decomposition of the diethylester **13**.



Scheme 3: a) MeOH/HCl, sodium hydrogencarbonate; b) diethyl chlorophosphite, chloroform, 62%;
c) H₂, Pd/C, MeOH, 95%.

For the synthesis of the phosphoramidate derivatives **15** (scheme 4) the crude amines **11** were treated with dibenzyl phosphite in tetrachloromethane.^{35,36}



Scheme 4: a) dibenzyl phosphite, tetrachloromethane, 63%; b) H₂, Pd/C, MeOH, 95%

Deprotection of the dibenzylphosphoryl groups³⁷ and the benzyloxycarbonyl amino group with hydrogen and palladium on charcoal afforded the target compounds **15** as the phosphoramidates. Biochemical properties of the synthesized compounds will be reported elsewhere.³⁸

EXPERIMENTAL

Solvents were purified in the usual way. Water sensitive reactions were carried out in flame dried glassware under argon. Thin layer chromatography: Merck precoated tlc plates, silica gel 60; column chromatography: Kieselgel 60 (Merck, 40-63 μm). Optical rotations. Perkin-Elmer polarimeter P 241. ^1H NMR: Bruker AM-250, Bruker AM-400, ^{13}C NMR: Bruker AM-250, ^{31}P NMR: Bruker AMX-300. Mass spectrometry: A. E. I. MS-30 and MS-50, ion source 180°C , FAB: Kratos Concept 1H, matrix = m-nitrobenzoic acid. Elemental analyses were performed at the Institute of Organic Chemistry and Biochemistry, Bonn, Microanalytical Department.

2-N-benzyloxycarbonyl-3-N-tert.-butoxycarbonyl-2 (S),3-diaminopropanoic acid (6)

The 2-N-Benzyloxycarbonyl-2 (S),3-diaminopropanoic acid (**5**) (5.0 g, 20.98 mmol) was suspended in dioxane/water (150 ml, 1 : 1 v/v) and triethylamine (4.19 ml, 31.48 mmol) was added. The resulting white suspension was cooled to 0°C and a solution of di-tert.-butyl dicarbonate (5.4 g, 2.98 mmol) in dioxane (50 ml) was added dropwise over 30 min. The mixture was allowed to warm to r. t. and stirred for 6 h at which time TLC analysis showed the reaction to be complete. The solvent was evaporated in vacuo to half its original volume, cooled in an ice-water bath, acidified to pH 5-6 by the slow addition of 0.1 N citric acid (50 ml), and then extracted with dichloromethane (3 x 100 ml). The combined organic extracts were dried over MgSO_4 , filtered and evaporated under reduced pressure to afford crude **6** which was crystallized from ethyl acetate/ether/petroleum ether to give pure **6**, yield: 6.03 g (85%), m. p. $144\text{--}146^\circ\text{C}$, R_f (n-butanol/AcOH/pyridine/water = 4 : 1 : 1 : 2) = 0.76).

^1H NMR (250 MHz, CDCl_3): δ = 1.40 (s, br, 9 H, $\text{C}(\text{CH}_3)_3$); 3.54 (m, 2 H, $\text{CH}_2\text{-CH}$); 4.36 (m, 1 H, CH-CH_2); 5.12 (s, 2 H, $\text{CH}_2\text{-C}_6\text{H}_5$); 5.75 (m, 1 H, NH-Boc); 6.19 (d, br, $J = 7.8$ Hz, 1 H, NHZ); 6.43 (s, br, 1 H, COOH); 7.34 (m, 5 H, C_6H_5).

^{13}C NMR (62.89 MHz, CDCl_3): δ = 28.48, 28.68, 28.89 ($\text{C}(\text{CH}_3)_3$); 46.27 ($\text{CH}_2\text{-NH}$); 55.49 ($\text{C}(\text{CH}_3)_3$); 67.64 (CH-COOH); 80.67 ($\text{O-CH}_2\text{-C}_6\text{H}_5$); 128.6, 129.10 (C_6H_5); 130.71, 136.18 (C_6H_5); 157.57 ($\text{NH-COO-C}(\text{CH}_3)_3$); 159.92 ($\text{NH-COO-CH}_2\text{-C}_6\text{H}_5$); 173.98 (COOH).

Analysis: $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_6$ (338.147) calcd. (%): C 56.78, H 6.56, N 8.30; found (%): C 56.67, H 6.47, N 8.30; MS (FAB-MS): $\text{C}_{16}\text{H}_{23}\text{N}_2\text{O}_6$ [$\text{M}+\text{H}$] $^+$, calcd.: $m/z = 339.155$, found: $m/z = 339.10$.

2-N-benzyloxycarbonyl-3-N-tert.-butoxycarbonyl-2 (S),3-diaminopropanoic acid-N-methoxy-N-methyl-carboxamide (7)

The protected diaminopropanoic acid **6** (3.59 g, 10.56 mmol) and triethylamine (2.8 ml, 21.25 mmol) were dissolved under argon atmosphere in THF (100 ml) and cooled to -23°C . Ethyl chloroformate (1.1 ml, 11.79 mmol) was added dropwise and the solution was stirred for 15 min at -23°C followed by addition of N, O-dimethylhydroxylamine hydrochloride (2.1 g, 21.25 mmol) and triethylamine (2.8 ml, 21.25 mmol). After 4 h the reaction was complete according to tlc. The precipitate was filtered off and washed with cold THF. The combined filtrates were dried over MgSO_4 , filtered and evaporated in vacuo affording a yellow oil which was

chromatographed on silica gel (petrolether/ethyl acetate 2 : 1, $R_f = 0.32$), yielded 2.47 g (61%) as a colourless oil.

^1H NMR (250 MHz, CDCl_3): $\delta = 1.39$ (s, br, 9 H, $\text{C}(\text{CH}_3)_3$); 3.19 (s, 3 H, N-CH_3); 3.32-3.59 (m, br, 2 H, $\text{CH}_2\text{-CH}$); 3.76 (s, 3 H, O-CH_3); 4.76 (m, 1 H, CH-CH_2); 4.86 (m, 1 H, NH-Boc); 5.08 (s, 2 H, $\text{CH}_2\text{-C}_6\text{H}_5$); 5.85 (d, br, $J = 8.2$ Hz, 1 H, NHZ); 7.33 (m, 5 H, C_6H_5).

^{13}C NMR (62.89 MHz, CDCl_3): $\delta = 21.68, 26.75, 28.94$ ($\text{C}(\text{CH}_3)_3$); 33.09 (N-CH_3); 42.67 ($\text{CH}_2\text{-CH}$); 52.44 ($\text{C}(\text{CH}_3)_3$); 58.70 (HNCH); 67.60 (O-CH_3); 80.27 ($\text{O-CH}_2\text{-C}_6\text{H}_5$); 128.71, 129.15 (C_6H_5), 131.72, 136.88 (C_6H_5); 156.65 ($\text{COO-C}(\text{CH}_3)_3$); 171.02 (NH-CO); 171.81 ($\text{COO-CH}_2\text{-C}_6\text{H}_5$).

Analysis: $\text{C}_{18}\text{H}_{27}\text{N}_3\text{O}_6 \times 0.15 \text{ H}_2\text{O}$ (383.89): calcd. (%): C 56.66, H 7.13, N 11.02; found (%): C 56.27, H 7.08, N 10.94; MS (HR-MS): $\text{C}_{16}\text{H}_{23}\text{N}_2\text{O}_6$ $[\text{M}+\text{H}]^+$, calcd.: $m/z = 382.1987$, found: $m/z = 382.1992$.

2-N-benzyloxycarbonyl-1-tert.-butoxycarbonyl-1,2 (S)-diamino-tridecan-3-ol (9)

The amide **7** (3.6 g, 9.5 mmol) was dissolved under argon in 100 ml absolute THF and cooled to -50°C . To this mixture 47.5 ml of 1 M solution of decylmagnesium bromide in diethyl ether (47.5 mmol) was added dropwise. After the reaction was completed (tlc analysis, petroleum ether/ethyl acetate 5 : 1), the mixture was poured onto 50 ml of 1 M NaH_2PO_4 while stirring vigorously, and extracted with ethyl acetate (3 x 50 ml). The combined organic extracts were washed subsequently with 1 M NaH_2PO_4 (1 x 30 ml) and brine (1 x 30 ml), dried over MgSO_4 , filtered and evaporated in vacuo. The residue was dissolved in 100 ml isopropyl alcohol and cooled to 0°C , sodium borohydride (1.07 g, 28.5 mmol) was added and the mixture was stirred for 12 h at 0°C . The reaction was quenched by dropwise addition of 120 ml 1 N HCl. The resulting solution was extracted with ethyl acetate (3 x 50 ml) and the combined organic phases were washed with saturated NaHCO_3 -solution (2 x 30 ml) and brine (1 x 30 ml), dried over MgSO_4 , filtered and evaporated in vacuo. The residue was chromatographed on silica gel (petrol ether/ethyl acetate 5 : 1, $R_f = 0.32$) yielding 2.03 g (46%) of the diastereomeric alcohols **9** as white powder (m. p. 186°C). As indicated by the integral ratio of the two signals of the 3-OH group in the ^1H NMR spectrum, **9** occurs as 3 : 1 mixture of the diastereomers.

^1H NMR (400 MHz, CDCl_3): $\delta = 0.85$ (t, $J = 6.6$ Hz, 3 H, CH_3); 1.12-1.56 (m, 27 H, Alkyl- CH_2 , $\text{C}(\text{CH}_3)_3$); 3.05 and 3.16 (d, br, $J = 6$ Hz, 1 H, diastereomeric OH, integration ratio 3 : 1); 3.32 (m, 1 H, CH-NH); 3.46-3.57 (m, 2 H, $\text{CH}_2\text{-CH}$); 3.62 (m, 1 H, CH-OH); 4.99 (m, 1 H, $\text{NHCOOC}(\text{CH}_3)_3$), 5.06 (s, 2 H, $\text{O-CH}_2\text{-C}_6\text{H}_5$); 5.33 (d, br., $J = 8.4$ Hz, 1 H, $\text{NHCOOCH}_2\text{C}_6\text{H}_5$); 7.31 (m, 5 H, C_6H_5).

^{13}C NMR (62.89 MHz, CDCl_3), values from the major diastereomer: $\delta = 14.78$ (CH_3); 19.21, 21.95, 21.45, 22.39, 22.58, 23.34, 26.70, 28.97, 29.99, 30.27, 32.57 ($\text{C}(\text{CH}_3)_3$, Alkyl- CH_2); 40.87 ($\text{CH}_2\text{-CHOH}$); 53.64 ($\text{C}(\text{CH}_3)_3$); 56.52 (CH-NH); 67.54 (CH-OH); 72.93 ($\text{CH}_2\text{-NH}$); 80.75 (O-CH_2), 129.20 (C_6H_5); 130.21, 131.62, 137.02 (C_6H_5); 157.09 ($\text{NHCOOC}(\text{CH}_3)_3$); 170.39 ($\text{NHCOOCH}_2\text{C}_6\text{H}_5$).

Analysis: $\text{C}_{26}\text{H}_{44}\text{N}_2\text{O}_5 \times 0.15 \text{ H}_2\text{O}$ (467.026): calcd. (%): C 66.89, H 9.49, N 5.99; found (%): C 67.20, H 9.76, N 5.62; MS (FAB-MS): $\text{C}_{26}\text{H}_{45}\text{N}_2\text{O}_5$ $[\text{M}+\text{H}]^+$, calcd.: $m/z = 465.332$, found: $m/z = 465.20$.

4-tert.-Butoxycarbonylaminoethyl-5-decyl-oxazolidin-2-one (10)

The diprotected amino alcohols **9** (150 mg, 0.3 mmol) were dissolved in methanol and palladium on charcoal (10%; caution: pyrophoric) was added. The mixture was stirred under 1 atm of H₂ for 12 h. After the reaction was completed (tlc analysis petrol ether/ethyl acetate 5 : 1), the catalyst was filtered off and the methanol was removed under reduced pressure to yield 95 mg (95%) of the deprotected amines as colourless oil. The crude amino alcohols were dissolved in dichloromethane (10 ml/mmol) and triphosgene (24.7 mg, 0.08 mmol) and triethylamine (131 μ l, 0.83 mmol) were added. After 1 h the reaction mixture was hydrolyzed with water. The organic layer was separated and washed successively with saturated sodium hydrogencarbonate solution and brine, dried over MgSO₄, filtered off and evaporated in vacuo. The residue was chromatographed on silica gel with petrol ether/ethyl acetate = 1 : 1 as eluent. **10** was obtained as a colourless oil, R_f = 0.42, yield = 68 mg (67 %).

¹H NMR (400 MHz, CDCl₃): δ = 0.81 (t, J = 6.89 Hz, 3 H, CH₃); 1.12-1.42 (m, 25 H, Alkyl-CH₂, C(CH₃)₃); 1.81-2.13 (m, 2 H, CH₂-CH); 3.34 dd, J = 8.39 Hz, J = 9.35 Hz, 1 H, CH₂-CHN); 3.37 (dd, J = 8.21 Hz, J = 9.34 Hz, 1 H, CH₂-CHN); 3.62 (ddd, J = 8.40 Hz, J = 3.6 Hz, J_{4,5} = 7.92 Hz, 1 H, 4-H, CH-N), values from the *erythro*-diastereomer; 3.81 (m, 1 H, CH-O); 4.69 (m, br, 1 H, NH).

O,O-Diethyl-N-[2 (S)-benzyloxycarbonylamino-3-hydroxy-tridecyl]-phosphoramidate (12)

The diastereomeric amino alcohols **9** (452 mg, 0.967 mmol) were dissolved in methanol saturated with hydrogen chloride (20 ml) and stirred for 12 h at ambient temperature. After the reaction was complete (tlc analysis, petrol ether/ethyl acetate 5 : 1), the aqueous solution was cooled to 0°C on an ice-water bath and neutralized with a saturated NaHCO₃ solution to pH = 7-8. The crude amines **11** were extracted with dichloromethane (3 x 10 ml) and the combined organic extracts were dried over MgSO₄, filtered and concentrated under reduced pressure to give 364 mg of crude amines **11** as white powder.

For the further reaction, the crude amines **11** (364 mg, 0.93 mmol) and triethylamine (125 μ l, 0.95 mmol) were dissolved in 30 ml chloroform and cooled to 0°C. To this mixture 153 μ l (0.96 mmol) of phosphoric acid diethylester chloride were added dropwise and stirring was continued for 12 h at ambient temperature. Volatile components were distilled off and the remaining residue was chromatographed on silica gel with dichloromethane/methanol = 30 : 1 as eluent. The phosphoramidates **12** were obtained as white powder, m. p. 186°C, R_f (dichloromethane/methanol = 30 : 1) = 0.42, yield = 312 mg (65.6%) as 3 : 1 mixture of diastereomers. The diastereomeric ratio was estimated by the integration ratio of the 3-OH signal.

¹H NMR (400 MHz, CDCl₃): δ = 0.86 (t, J = 6.54 Hz, 3 H, CH₃); 1.17-1.32 (m, 24 H, Alkyl-CH₂, P(O-CH₂CH₃)₂); 1.41-1.53 (m, 2 H, CH₂-CH); 2.94 and 3.02 (d, br, J = 5.8 Hz, 1 H, diastereomeric OH, integration ratio 3 : 1); 3.19-3.69 (m, 5 H, CH₂-NH, CH-NH, CH-OH, NHCOOC(CH₃)₃); 3.98 (q, J = 7 Hz, 4 H, P(OCH₂CH₃)₂); 5.07 (s, 2 H, CH₂-C₆H₅); 5.80 (d, br, J = 8 Hz, 1 H, NHCOOCH₂C₆H₅); 7.32 (m, 5 H, C₆H₅).

¹³C NMR (62.89 MHz, CDCl₃), values from the major diastereomer: δ = 14.14 (CH₃); 16.05, 16.16 (P(OCH₂CH₃)₂); 22.70, 26.15, 29.38, 29.66, 31.92, 33.07, 33.14, 33.91 (Alkyl-CH₂); 41.21 (CH₂CHOH); 55.32 (CHNH); 62.53, 62.76 (P(OCH₂CH₃)₂); 67.11 (CHOH); 72.38 (CH₂NH); 82.25 (OCH₂C₆H₅); 128.06, 128.35, 132.56, 136.59 (C₆H₅); 156.55 (COOCH₂C₆H₅).

^{31}P NMR (121.49 MHz, CDCl_3): $\delta = 9.17$.

Analysis: $\text{C}_{25}\text{H}_{45}\text{N}_2\text{O}_6\text{P}$ (500.301): calcd. (%): C 59.98, H 9.06, N 5.60; found (%): C 60.02, H 9.25, N 5.72; MS (FAB-MS): $\text{C}_{25}\text{H}_{46}\text{N}_2\text{O}_6\text{P}$ $[\text{M}+\text{H}]^+$, calcd.: $m/z = 501.301$, found: $m/z = 501.42$.

O,O-Diethyl-N-[2(S)-amino-3-hydroxy-tridecyl]-phosphoramidate (13)

The phosphoramidates **12** (200 mg, 0.4 mmol) were dissolved in methanol and palladium on charcoal (10%; caution: pyrophoric) was added. The mixture was stirred under 1 atm of H_2 for 12 h. After the reaction was completed (tlc analysis dichloromethane/methanol 30 : 1), the catalyst was filtered off and the methanol was removed under reduced pressure to yield 135 mg (95%) of **13** as colourless oil (R_f (dichloromethane/methanol/1% ammonia 10 : 1) = 0.23). As indicated by the integral ratio of the two signals of the 3-OH group in the ^1H NMR spectrum, **13** occurs as 3 : 1 mixture of the diastereomers.

^1H NMR (400 MHz, CDCl_3): $\delta = 0.85$ (t, $J = 7$ Hz, 3 H, CH_3); 1.18-1.26 (m, 16 H, Alkyl- CH_2); 1.29 (t, $J = 7.1$ Hz, 6 H, $\text{P}(\text{OCH}_2\text{CH}_3)_2$); 1.41-1.46 (m, 2 H, $\text{CH}_2\text{-CHOH}$); 2.89 and 3.04 (d, br, $J = 6.2$ Hz, 1 H, diastereomeric OH, integration ratio 3 : 1); 3.10-3.47 (m, 4 H, $\text{CH}_2\text{-NH}$, NH_2); 3.57-3.85 (m, 2 H, CH-NH , CH-OH); 4.04 (dq, $J = 7.1$ Hz, $^3J_{\text{HP}} = 2.75$ Hz, $\text{P}(\text{OCH}_2\text{CH}_3)_2$).

^{13}C NMR (62.89 MHz, CDCl_3), values from the major diastereomer: $\delta = 14.32$ (CH_3); 16.26, 16.32 ($\text{P}(\text{OCH}_2\text{CH}_3)_2$); 22.73, 24.64, 25.92, 26.32, 29.55, 31.8, 33.07, 33.53 (Alkyl- CH_2); 42.48 (CH_2CHOH); 56.42 (CHNH_2); 62.68, 62.83 ($\text{P}(\text{OCH}_2\text{CH}_3)_2$); 67.31 (CHOH); 72.56 (CH_2NH).

^{31}P NMR (121.49 MHz, CDCl_3): $\delta = 9.21$.

Analysis: $\text{C}_{17}\text{H}_{39}\text{N}_2\text{O}_4\text{P}$ (366.480): calcd. (%): C 55.72, H 10.73, N 7.64; found (%): C 55.36, H 10.44, N 7.31; MS (FAB-MS): $\text{C}_{17}\text{H}_{40}\text{N}_2\text{O}_4\text{P}$ $[\text{M}+\text{H}]^+$, calcd.: $m/z = 367.47$, found: $m/z = 367.40$.

O,O-Dibenzyl-N-[2(S)-benzyloxycarbonylamino-3-hydroxy-tridecyl]-phosphoramidate (14)

The diastereomeric amino alcohols **9** (460 mg, 0.98 mmol) were dissolved in methanol saturated with hydrogen chloride (20 ml) and stirred for 12 h at ambient temperature. After the reaction was complete (tlc analysis, petrol ether/ethyl acetate 4 : 1), the aqueous solution was cooled to 0°C on an ice-water bath and neutralized with a saturated NaHCO_3 solution to $\text{pH} = 7-8$. The crude amines **11** were extracted with dichloromethane (3 x 10 ml) and the combined organic extracts were dried over MgSO_4 , filtered and concentrated under reduced pressure to give 380 mg of crude amines **11** as white powder.

For the further reaction, the crude amines **11** (380 mg, 0.96 mmol) were dissolved under argon in 5ml tetrachloromethane. To this solution 115 μl (0.52 mmol) of dibenzylphosphite were added dropwise and the mixture was stirred for 12 h at ambient temperature. The solvent was concentrated under reduced pressure and the remaining residue was chromatographed on silica gel with dichloromethane/methanol 40 : 1 to yield 442 mg (68%) of **14** as white powder, R_f (dichloromethane/methanol 40 : 1) = 0.27.

^1H NMR (400 MHz, CDCl_3): $\delta = 0.86$ (t, $J = 6.43$ Hz, 3 H, CH_3); 1.14-1.48 (m, 18 H, Alkyl- CH_2); 2.87 and 2.98 (d, br, 1 H, diastereomeric OH, integration ratio 3 : 1); 3.02-3.30 (m, 3 H, NH , CH_2NH); 3.41-3.68 (m, 2 H, CH-NH , CH-OH); 4.96 (s, 2 H, $\text{O-CH}_2\text{-C}_6\text{H}_5$); 5.00 (s, 2 H, $\text{O-CH}_2\text{-C}_6\text{H}_5$); 5.04 (s, 2 H, $\text{COO-CH}_2\text{-C}_6\text{H}_5$);

5.45 (d, br, $J = 7.72$ Hz, 1 H, $\text{NHCOOCH}_2\text{C}_6\text{H}_5$); 7.29 (s, 10 H, $\text{P}(\text{OCH}_2\text{-C}_6\text{H}_5)_2$); 7.32 (m, 5 H, $\text{COOCH}_2\text{C}_6\text{H}_5$).

^{13}C NMR (62.89 MHz, CDCl_3), values from the major diastereomer: $\delta = 14.34$ (CH_3); 22.70, 24.6, 26.15, 29.38, 29.66, 31.92, 33.14, 33.91 (Alkyl- CH_2); 41.21 (CH_2CHOH); 55.32 (CHNH); 67.11 (CHOH); 72.38 (CH_2NH); 82.25 ($\text{OCH}_2\text{C}_6\text{H}_5$); 84.35, 85.21 ($\text{P}(\text{OCH}_2\text{C}_6\text{H}_5)_2$); 128.12, 128.35, 128.42, 128.47, 129.43, 130.23, 131.91, 132.12, 132.56, 135.67, 136.34, 136.59 ($\text{P}(\text{OCH}_2\text{C}_6\text{H}_5)_2$, $\text{COOCH}_2\text{C}_6\text{H}_5$); 156.55 ($\text{COOCH}_2\text{C}_6\text{H}_5$).

^{31}P NMR (121.49 MHz, CDCl_3): $\delta = 9.73$.

Analysis: $\text{C}_{35}\text{H}_{49}\text{N}_2\text{O}_6\text{P}$ (624.756): calcd. (%): C 67.29, H 7.91, N 4.48; found (%): C 67.29, H 8.02, N 7.89;

MS (FAB-MS): $\text{C}_{35}\text{H}_{50}\text{N}_2\text{O}_6\text{P}$ $[\text{M}+\text{H}]^+$, calcd.: $m/z = 625.34$, found: $m/z = 625.40$.

N-[2 (S)-amino-3-hydroxy-tridecyl]-phosphoramidate (15)

The phosphoramidate dibenzylester **14** (256 mg, 0.41 mmol) was dissolved in methanol and palladium on charcoal (10%; caution = pyrophoric) was added. The reaction mixture was stirred under 1 atm of H_2 for 12 h. After the reaction was completed (tlc analysis, dichloromethane/methanol 30 : 1), the catalyst was filtered off and the methanol was removed under reduced pressure to yield 122 mg (96%) of **15** as colourless oil (R_f (chloroform/methanol/water 60 : 35 : 8) = 0.29) as 3 : 1 mixture of diastereomers. The diastereomeric ratio was estimated by the integration ratio of the 3-OH signal.

^1H NMR (400 MHz, CDCl_3): $\delta = 0.81$ (t, $J = 6.23$ Hz, 3 H, CH_3); 1.16-1.52 (m, 16 H, Alkyl- CH_2); 2.50 and 2.68 (d, br, $J = 5.7$ Hz, 1 H, diastereomeric OH, integration ratio 3 : 1); 2.63-2.96 (m, 2 H, $\text{CH}_2\text{-OH}$); 3.01-3.32 (m, 1 H, CH-NH_2); 3.43-3.68 (m, 2 H, $\text{CH}_2\text{-NH}$); 3.82-3.98 (m, 1 H, CH-OH); 5.4-6.6 (s, br, 4 H, $\text{P}(\text{OH})_2$, NH_2).

^{13}C NMR (62.89 MHz, CDCl_3), values from the major diastereomer: $\delta = 14.23$ (CH_3); 21.72, 23.6, 25.35, 28.38, 28.76, 30.72, 33.54, 33.81 (Alkyl- CH_2); 42.21 (CH_2CHOH); 55.62 (CHNH_2); 67.51 (CHOH); 73.45 (CH_2NH).

Analysis: $\text{C}_{13}\text{H}_{31}\text{N}_2\text{O}_4\text{P} \times 0.3 \text{ H}_2\text{O}$ (315.426) calcd. (%): C 49.45, H 10.09, N 8.87; found (%): C 49.12, H 10.12, N 8.67 MS (FAB-MS): $\text{C}_{13}\text{H}_{32}\text{N}_2\text{O}_4\text{P}$ $[\text{M}+\text{H}]^+$, calcd.: $m/z = 311.209$, found: $m/z = 311.30$.

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REFERENCES

1. Sweeley, C. C., in "Biochemistry of Lipids and Membranes", Vance D. E., Vance, J. E., Eds., Benjamin/Cummings Publishing Co., Menlo Park, CA, 1985.
2. a) Hakomori, S., *Trends Biochem. Sci.* **1984**, *9*, 453-458;
b) Hakomori, S., Igarashi, Y., *Adv. Lipid Res.* **1993**, *25*, 147-162;
c) Hakomori, *Biochem. Soc. Trans.* **1993**, *21* (Pt 3), 583-595.
3. Rother, J., van Echten, G., Schwarzmann, G., Sandhoff, K., *Biochem. Biophys. Res. Commun.* **1992**, *189*, 14-20.

4. a) Hannun, Y. A., Loomis, C. R., Merrill, A. H., Bell, R. M., *J. Biol. Chem.* **1986**, *261*, 12604-12609;
b) Hannun, Y. A., Bell, R. M., *Science* **1987**, *235*, 670-674.
5. a) Hannun, Y. A., Bell, R. M., *Science* **1989**, *243*, 500-507;
b) Merrill, A. H., Stevens, V. L., *Biochim. Biophys. Acta* **1986**, *1010*, 131-139.
6. Zhang, H., Desai, N. N., Olivera, A., Seki, T., Brooker, G., Spiegel, S., *J. Cell Biol.* **1991**, *114*, 155-167.
7. a) Hirschberg, C. B., Kisic, A., Schroepfer, G. J., *J. Biol. Chem.* **1970**, *254*, 3084-3090;
b) Stoffel, W., Hellenbroich, B., Heiman, G., *Hoppe-Seyler's Z. Physiol. Chem.* **1973**, *354*, 1311-1316;
c) Keenan, R. W., Okabe, K., *Biochemistry* **1968**, *7*, 2696-2701;
d) Stoffel, W., Assman, G., Binczeck, E., *Hoppe-Seyler's Z. Physiol. Chem.* **1970**, *351*, 635-642;
e) Louie, D. D., Kisic, A., Schroepfer, G. J., *J. Biol. Chem.* **1976**, *251*, 4557-4564.
8. Buehrer, B. M., Bell, R. M., *Adv. Lipid Res.* **1993**, *26*, 59-67.
9. a) Stoffel, W., Bauer, E., Stahl, J., *Hoppe-Seyler's Z. Physiol. Chem.* **1973**, *355*, 61-74.
b) Stoffel, W., Sticht, G., Le Kim, D., *Hoppe-Seyler's Z. Physiol. Chem.* **1968**, *349*, 1745-1748;
c) Shimojo, T., Akino, T., Miura, Y., Schroepfer, G. J., *Biochim. Biophys. Acta* **1976**, *431*, 433-446;
d) Van Veldhoven, P. P., Mannaerts, G. P., *Adv. Lipid Res.* **1993**, *26*, 69-98.
10. a) Van Veldhoven, P. P., Mannaerts, G. P., *Biochem. J.* **1994**, *299*, 597-601;
b) Van Veldhoven, P. P., De Ceuster, P., Rozenberg, R., Mannaerts, G. P., de Hoffmann, E., *FEBS Lett.* **1994**, *350*, 91-95.
11. Desai, N. N., Zhang, H., Olivera, A., Mattie, M. E., Spiegel, S., *J. Biol. Chem.* **1992**, *267*, 23122-23128.
12. Su, Y., Rosenthal, D., Samulson, M., Spiegel, S., *J. Biol. Chem.* **1994**, *269*, 16512-16517.
13. Bornfeldt, K. E., Graves, L. M., Raines, E. W., Igarashi, Y., Wayman, G., Yamamura, S., Yatomi, Y., Sidhu, J. S., Krebs, E. G., Hakomori, S. I., Ross, R., *J. Cell Biol.* **1995**, *130*, 193-206.
14. Goodemote, K. A., Matties, M. E., Berger, A., Spiegel, S., *J. Biol. Chem.* **1995**, *270*, 10272-10277.
15. Gosh, T. K., Bian, J., Gill, D. L., *Science* **1990**, *248*, 1653-1654.
16. Spiegel, S., Olivera, A., Zhang, H., Thompson, E. W., Su, Y., Berger, A., *Breast Cancer Res. and Treat.* **1994**, *31*, 337-348
17. a) Sadahira, Y., Ruan, F., Hakomori, S., Igarashi, Y., *Proc. Natl. Acad. Sci. U.S.A.* **1992**, *89*, 9686-9690;
b) Ruan, F., Sadahira, Y., Hakomori, S., Igarashi, Y., *Bioorg. Med. Chem. Lett.* **1992**, *2*, 973-978.
18. Bornfeldt, K. E., Graves, L. M., Raines, E. W., Igarashi, Y., Wayman, G., Yamamura, S., Yatomi, Y., Sidhu, J. S., Krebs, E. G., Hakomori, S., Ross, R., *J. Cell Biol.* **1995**, *130*, 193-206.
19. Sem, D. S., Cleland, W. W., *Biochemistry* **1991**, *30*, 4978-4984.
20. recent examples: a) Knight, W. B., Cleland, W. W., *Biochemistry* **1989**, *28*, 5728-5734;
b) Sheffer-Dee-Noor, S., Baasov, T., *Bioorg. Med. Chem. Lett.* **1993**, *3*, 1615-1618;
c) Rajagopalan, K., Chavan, A. J., Haley, B. E., Watt, D. S., *J. Biol. Chem.* **1993**, *268*, 14230-14238.
21. Schick, A., Kolter, T., Giannis, A., Sandhoff, K., *Tetrahedron* **1995**, *51*, 11207-11218.
22. a) Egbertson, M. S., Homnick, C. F., Hartmann, G. D., *Synthetic. Comm.* **1993**, *23*, 703-709;
b) Waki, M., Kitajima, Y., Izumiya, N., *Synthesis* **1981**, 266-268;
c) Itoh, M., Hagiwara, D., Kamiya, T., *Bull. Chem. Soc. Jpn.* **1977**, *50*, 718-722.
23. König, W., Geiger, R. *Chem. Ber.* **1970**, *103*, 788-793.
24. Nahm, S., Weinreb, S. M., *Tetrahedron Lett.* **1981**, *22*, 3815-3823.
25. a) Radunz, H. E., Devant, R. M., Eiermann, V., *Liebigs Ann. Chem.* **1988**, 1103-1105;
b) Jurczak, J., Pikul, S., Bauer, T., *Tetrahedron* **1986**, *42*, 447-453;
26. Dufuor, M.-N., Jouin, P., Poncet, J., Pantaloni, A., Castro, B., *J. Chem. Soc. Perkin Trans. I* **1986**, 1895-1902.
27. Kano, S., Yokomatsu, T., Iwasawa, H., Shibuya, S., *Chem. Pharm. Bull.* **1988**, *36*, 3296-3304.
28. a) Boutin, R. H., Rapoport, H., *J. Org. Chem.* **1985**, *51*, 5320-5326;

- b) Cups, T. L., Boutin, R. H., Rapoport, H., *J. Org. Chem.* **1985**, *50*, 3972-3984.
29. a) Miyashita, M., Yoshikoshi, A., Grieco, P. A., *J. Org. Chem.* **1979**, *42*, 3772-3774;
b) Sterzycki, R., *Synthesis* **1979**, 724-724.
30. Eckert, H., Forster, B., *Angew. Chem.* **1987**, *99*, 922-934.
31. Wolf, J. P., Pfander, H., *Helv. Chim. Acta* **1986**, *69*, 918-924.
32. a) Dale, J. A., Mosher, H. S., *J. Am. Chem. Soc.* **1973**, *95*, 512-518;
b) Dale, J. A., Dull, D. L., Mosher, H. S., *J. Org. Chem.* **1969**, *34*, 2543-2549.
33. a) Li, S. O., Eakin, R. E., *J. Am. Chem. Soc.* **1955**, *77*, 1866-1872;
b) Zhao, Y.-F., Ji, G.-J., Xi, S.-K., Tang, H.-G., Song, A.-T., Wie, S.-Z., *Phosphorus and Sulfur* **1983**, *18*, 155-158.
34. Tong, Z. S., *Acta. Chim. Sinica* **1981**, *39*, 69-73.
35. a) Zhao, Y.-F., Xi, S.-K., Song, A.-T., Ji, G.-J., *J. Org. Chem.* **1984**, *49*, 4549-4553;
b) Cosmatos, A., Photaki, I., Zervas, L., *Chem. Ber.* **1961**, *94*, 2644-2652.
36. Wolfrom, M. L., Conigliaro, P. J., Soltes, E. J., *J. Org. Chem.* **1967**, *32*, 653-659.
37. Petrov, K. A., Bliznjuk, N. K., Lysenko, T. N., *Z. Obsc. Chim.* **1960**, *30*, 1964-1969.
38. Schick, A., van Echten-Deckert, G., Sandhoff, K., manuscript in preparation.

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